

## Pyrrolo[3,2-*c*]pyridine Derivatives as Inhibitors of Platelet Aggregation

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**Abstract**—A series of pyrrolo[3,2-*c*]pyridines, isosteres of the antithrombotic drug ticlopidine, has been synthesized and evaluated in vitro for the ability to inhibit aggregation of human platelet-rich plasma induced by adenosin 5'-diphosphate (ADP). Structure–activity relationships showed their antiplatelet effects to be related to the lipophilicity. © 2000 Elsevier Science Ltd. All rights reserved.

Platelet aggregation has an important role in the thrombotic events associated to relevant cardiovascular diseases.<sup>1</sup> As a part of our ongoing research directed toward the development of novel antiplatelet agents, we synthesized a series of 4,5,6,7-tetrahydropyrrolo[3,2-*c*]pyridines and evaluated their effects on in vitro platelet aggregation. Pyrrolo[3,2-*c*]pyridine derivatives are isosteres of the currently used thieno[3,2-*c*]pyridine antithrombotic agents, among which ticlopidine (**1**), 5-[(2-chlorophenyl)methyl]-4,5,6,7-tetrahydrothieno[3,2-*c*]pyridine, and its derivatives (e.g. clopidogrel) have been proven to be effective in the treatment and/or prevention of platelet-dependent disorders,<sup>2</sup> including thrombotic stroke and suppression of platelet activation after coronary stenting<sup>3</sup> (Fig. 1).

Ticlopidine interferes with platelet membrane functions by inhibiting the binding of adenosine 5'-diphosphate (ADP) to its platelet receptors and subsequent platelet–platelet interactions. It also reduces deposition of platelets and fibrin on artificial surface, and prolongs the bleeding time.<sup>4</sup> The effects on platelet functions are irreversible for the life of platelet, as shown by ex vivo measurements of the platelet aggregation inhibition.<sup>5</sup>

We herein report the synthesis, the effects on in vitro ADP-induced platelet aggregation, and preliminary structure–activity relationships of a series of 4,5,6,7-tetrahydropyrrolo[3,2-*c*]pyridines.

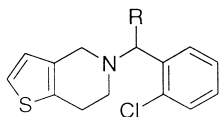
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### Synthesis

Tetrahydropyrrolo[3,2-*c*]pyridines (**2–11**) were synthesised in satisfactory yields as shown in Scheme 1. Compounds **2**, **3**, **5**, **6** and their N(1)-vinyl derivatives (e.g. **4**) were synthesised by applying the Trofimov reaction<sup>6</sup> to 1-benzyl-, 1-ethyl-, 2,5-dimethyl-, and 1,2,5-trimethyl-piperidine-4-one oximes. Vilsmeier-Haack formylation of **6** and 1-vinyl-4,5,6,7-tetrahydro-5-benzylpyrrolo[3,2-*c*]pyridine provided compounds **7** and **4**, respectively. Pyrrolopyridines **9**, **10** and **11** were obtained, respectively, by condensation with ethanolamine and malononitrile, and by reduction with NaBH<sub>4</sub> of the formyl derivative **7**. The nitration of **6** under Mencke reaction conditions<sup>7</sup> gave the 2-nitro derivative **8**. The synthesis of compounds **2–4** and **10** will be fully described elsewhere, whereas the other derivatives tested in this study were prepared according to reported procedures.<sup>8</sup> As for diastereomeric 4,7-dimethyl substituted pyrrolopyridines **5–11**, NMR data indicated that piperidine ring has a semi-chair conformation with *trans* diequatorial 4- and 7-CH<sub>3</sub> groups.<sup>9</sup>

### Antiplatelet Effects

Pyrrolo[3,2-*c*]pyridine derivatives **2–11** were evaluated as platelet aggregation inhibitors by measuring their effect on the in vitro aggregation of human platelet-rich plasma (PRP) induced by ADP, by using a turbidimetric method.<sup>10</sup> The results are summarized in Table 1. With the exception of 2-nitro- (**8**) and 2-hydroxymethyl-



**Figure 1.** Structure of the antithrombotic agents ticlopidine (**1**, R = H) and clopidogrel (R = COOCH<sub>3</sub>).

(**11**) 4,5,7-trimethyl-4,5,6,7-tetrahydropyrrole[3,2-*c*]pyridine derivatives, all the test compounds appreciably inhibited the platelet aggregation at doses close to the IC<sub>50</sub> value of ticlopidine.

Within the limits of this screening, the closest analogue (**2**) of ticlopidine and the 2-formyl derivative **7** showed the highest activities. The aggregation tracing relative to the control revealed a typical biphasic curve. In general, the test compounds inhibited the second phase of in vitro aggregation, whereas the most active ones (e.g. compound **7**) partially inhibited also the primary wave.

Overall, pyrrolo[3,2-*c*]pyridines exhibit a moderate, but structure-dependent, inhibition of in vitro platelet activation, two of them (**2** and **7**) having an activity comparable to that of ticlopidine. Actually, ticlopidine itself is known to display weak platelet inhibitory effects when assayed in vitro at the active concentration attained in vivo.<sup>5</sup> Nevertheless, the apparent relationship between the in vitro antiplatelet effect and the in vivo antithrombotic potency, observed for other series of compounds (e.g. 3-carbamoylpiperidines),<sup>11</sup> prompt us to examine further pyrrolopyridine derivatives and to select candidates for in vivo and/or ex vivo activity measurements.

### Lipophilicity and Related Parameters

To detect the physicochemical factors possibly related to the inhibition of ADP-mediated platelet aggregation,

we undertook an examination of the partitioning behaviour, by calculating 1-octanol/water partition coefficients and measuring retention in RP-HPLC.

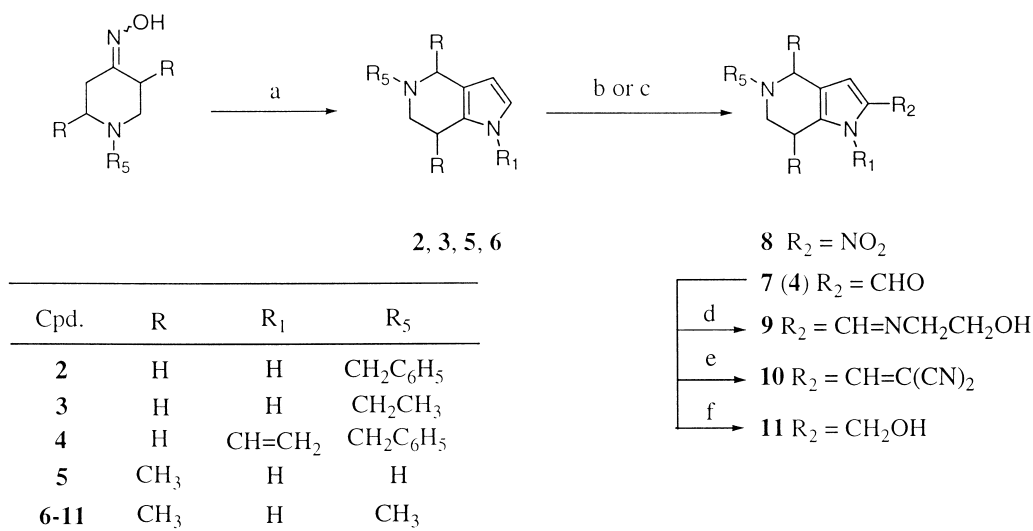
Log *P* values (Table 1), calculated by CLOG P software,<sup>12</sup> based on the fragmental method of Hasch and Leo,<sup>13</sup> indicated that the compounds under examination cover a range of about 4 log units. For a number of compounds, distribution coefficients (log *D*) at physiological pH (7.40), where piperidine nitrogen is mainly in the protonated form, were also measured using the conventional 'shake-flask' (SF) technique.<sup>14</sup>

RP-HPLC retention data were measured using a new silanol-deactivated octadecylsilane (ODS)<sup>15</sup> as the non-polar stationary phase, at 0.05-increments of the volume fraction of methanol in the aqueous mobile phase ( $\phi_{\text{MeOH}}$ ). In agreement with previous results,<sup>16,17</sup> the capacity factors (log *k'*) of pyrrolo[3,2-*c*]pyridines increased linearly ( $r^2 > 0.95$ ) with decreasing methanol concentration in the mobile phase in the range  $0.1 < \phi_{\text{MeOH}} < 0.7$ . Thus, by using the following linear relationship

$$\log k' = \log k'_w - s\phi$$

log *k'*<sub>w</sub> that is the logarithm of the capacity factor extrapolated to 100% water in the mobile phase, and *s* (slope), that is a constant for the solute-eluent combination, were calculated and reported in Table 1.

Other authors had observed that comparing between them the slope *s* and log *k'*<sub>w</sub> may help in unraveling differences in polarity and H-bonding (HB) properties within a given series of compounds.<sup>18–20</sup> With our compounds, the variations in log *k'*<sub>w</sub> values are more or less those expected from log *P* scale, whereas *s* parameter appeared to be more dependent on the HB capacity of the compounds. In fact, ticlopidine (**1**) and compound **4**, two net HB acceptors, have *s* value of ca. 1.4, whereas



**Scheme 1.** (a) C<sub>2</sub>H<sub>2</sub>, KOH, DMSO, 80–90 °C, 4–6 h, 40–50%; (b) **8**: Cu(NO<sub>3</sub>)<sub>2</sub>, Ac<sub>2</sub>O, rt 5 h, 47%; (c) **4**, **7**: POCl<sub>3</sub>, DMF, rt, 5 h, 78–80%; (d) NH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>OH, toluene, reflux, 5 h, 92%; (e) CH<sub>2</sub>(CN)<sub>2</sub>, EtOH, reflux, 3–5 h 95%; (f) NaBH<sub>4</sub>, EtOH, rt, 5 h, 84%.

**Table 1.** Antiplatelet activity and physicochemical data of 4,5,6,7-tetrahydropyrrolo[3,2-*c*]pyridine derivatives **1–11**

Compound	% Inhibition of platelet aggregation <sup>a</sup>	CLOG P <sup>b</sup>	Log D <sup>c</sup>	RP-HPLC retention data <sup>d</sup>	
				Log $k'_w$	<i>s</i>
Control	6.0 ± 1.1				
<b>1</b>	48.5 ± 5.9***	4.04	>3.00	1.99	1.41
<b>2</b>	46.5 ± 8.9***	2.24	1.59	1.66	4.12
<b>3</b>	23.7 ± 5.0**	1.20		0.60	5.58
<b>4</b>	38.7 ± 0.5***	3.08		1.29	1.42
<b>5</b>	26.0 ± 14.5	1.24	−1.08	0.88	4.40
<b>6</b>	28.5 ± 6.6**	1.91	−0.41	0.96	5.06
<b>7</b>	48.0 ± 7.3***	1.72	0.69	0.93	6.24
<b>8</b>	3.6 ± 1.6	2.13	1.14	0.97	4.77
<b>9</b>	12.2 ± 7.1	−0.05		<sup>f</sup>	
<b>10</b>	ND <sup>e</sup>	1.19		1.39	3.04
<b>11</b>	3.3 ± 3.3	0.87		1.02	7.44

<sup>a</sup>Platelet-rich plasma (PRP) was pre-incubated with the test compounds (250 μM) or with dimethylsulphoxide (0.5%, control) at 37 °C for 4 min. The inducer ADP (10 μM) was then added. Percent inhibition of aggregation is presented as means ± SEM (*n* = 4–6). \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.001; significantly different from the respective control value.

<sup>b</sup>Calculated log *P* values.<sup>12</sup>

<sup>c</sup>1-Octanol/water distribution coefficients measured at pH 7.40 (0.01 MOPS buffer).

<sup>d</sup>Capacity factors extrapolated at 100% water eluent (log  $k'_w$ ) and slope (*s*) determined on an octadecylsilane stationary phase.<sup>15</sup>

<sup>e</sup>ND = not determined, because of its low solubility.

<sup>f</sup>Due to its hydrophilicity, it was not retained even at highly polar mobile phase (>90% water).

all the others pyrrolopyridine derivatives have *s* < 3.0. The total HB donor capacity (see **5** and **6**) seems to depend essentially on the pyrrole NH, and marginally on the piperidine NH, as assessed by the Abraham's  $\alpha_2^H$  descriptors of HB donor capacity (0.41 and 0.06, respectively).<sup>21</sup> The maximum value of *s* parameter (ca. 7.4) observed for **11** is consistent with the dominance of conformations lacking intramolecular H-bond between pyrrole NH and OH group of hydroxymethyl substituent in 2 position.

Interestingly, for the congeners **7** (2-CHO) and **8** (2-NO<sub>2</sub>), the log  $k'_w$  values are almost identical, whereas the *s* values differ significantly (6.24 and 4.77, respectively). Moreover, the log  $k'_w$  and *s* values of compound **7** and the unsubstituted congener **6** are close, suggesting that polarity/polarizability and HB acceptor ability of the nitro substituent should negligibly affect the RP-HPLC retention behaviour. To gain insights into conformations and/or self-association properties of congeners **6**, **7** and **8**, we examined their FT-IR spectra in chloroformic solutions, at concentrations below 5 × 10<sup>−2</sup> M. The  $\nu_{NH}$  absorption at 3479 cm<sup>−1</sup> for compound **6** was assigned to the stretching of free pyrrole NH. A shift of the stretching vibration band  $\nu_{NH}$  to lower wavenumber (3445 cm<sup>−1</sup>,  $\Delta\nu_{NH}$  = 34 cm<sup>−1</sup>) for the 2-CHO congener (**7**) could account for an intramolecular HB, NH...O=C, or a dipolar alignment. A smaller  $\Delta\nu_{NH}$  (ca. 10 cm<sup>−1</sup>) found for the nitro derivative **8** revealed a weaker acceptor ability of NO<sub>2</sub> group and probably a lower contribution of the internally H-bonded conformations. A weaker and broader, but

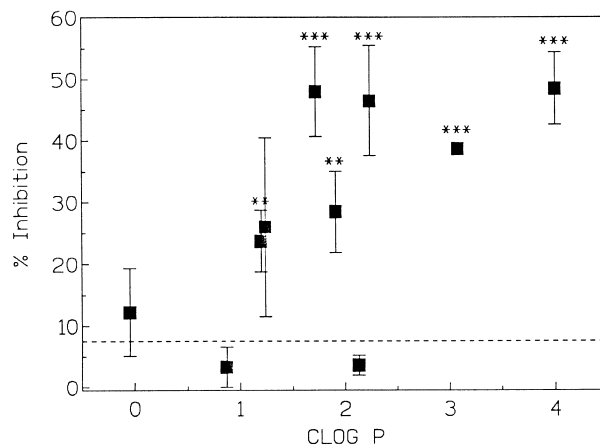
concentration-dependent, band at 3270 cm<sup>−1</sup>, assigned to  $\nu_{NH}$  vibration of intermolecular (self-associated) hydrogen-bonded species (NH...O=C), was observed in the IR spectrum of **7**, and not in that of the NO<sub>2</sub> congener **8**. This indicated that CHO group is more effective than NO<sub>2</sub> group in increasing the HB donor ability of the pyrrole NH. Most likely, the torsion angle of the nitro group is far from 0°, and this loss of coplanarity could result in a diminished electronic conjugation.<sup>22</sup>

### Lipophilicity–Activity Relations

A comparison of antiplatelet effects values with physicochemical parameters suggested trends of correlations. Lipophilicity appears to play a role in modulating the antiplatelet activity (Fig. 2).

In fact, with the exception of less active derivatives **8** and **11**, when lipophilicity increases activity increases as well, until a value of log *P* around 2 is reached. A vinyl group on pyrrole NH (**4**) is not beneficial, and the methyl groups in 4 and 7 positions should not exert any different effect from those accounted for by the lipophilicity. As far as the role of the 2-X substituents is concerned, our results indicated that, besides the additive contribution of the 2-X fragments to lipophilicity, which could explain the low activity of the most hydrophilic congeners **9** and **11**, other factors, especially the electronic and conformational ones, should be taken into account. Thus, the formyl congener (**7**), equiactive with ticlopidine despite its lower log *P* value, resulted significantly more active than the nitro congener (**8**). The *s* parameter from RP-HPLC and IR spectroscopy revealed that the formyl group (and not the nitro group) influences the ability of NH to act as a HB donor, suggesting a role of the pyrrole NH in modulating the inhibition of platelet aggregation.

In conclusion, our study showed 4,5,6,7-tetrahydropyrrolo[3,2-*c*]pyridines to be inhibitors of ADP-stimulated platelet aggregation in vitro, their activity being related to the lipophilicity. Actually, the importance of



**Figure 2.** Plot of inhibition of ADP-induced platelet aggregation against lipophilicity as assessed by CLOG *P* values. The dashed line represents the control inhibition.

lipophilicity in the antiplatelet activity had been demonstrated by others for different series of derivatives.<sup>23</sup> Our results highlight other properties, especially H-bonding, likely involved in the platelet aggregation inhibition of pyrrolo[3,2-*c*]pyridines, and stimulate a deeper examination of further isosteres and analogues of ticlopidine-related compounds.

### References and Notes

1. Majerus, W.; Broza, G. J.; Miletich, J. P.; Tollesfsen, D. M. In *The Goodman Gilman's Pharmacological Basis of Therapeutics*; Goodman, L. S.; Gilman, A. G.; Limbird, L. E., Eds.; McGraw-Hill: New York, 1996; pp 1341–1359.
2. Gardell, S. J. *Perspect. Drug Discov. and Des.* **1993**, *1*, 521.
3. Newmann, F. J.; Gawaz, M.; Dickfelt, T.; Wehinger, A.; Walter, H.; Blasini, R.; Schomig, A. *Ann. Intern. Med.* **1998**, *29*, 394.
4. Heptinstall, S.; May, J. A.; Glenn, J. R.; Sanderson, H. M.; Dickinson, J. P.; Wilcox, R. G. *Thromb. Haemostasis* **1995**, *74*, 1310.
5. Shrör, K. *Platelets* **1993**, *4*, 252.
6. Trofimov, B. A.; Mikhaleva, A. I. *Heterocycles* **1994**, *37*, 1193.
7. Menke, J. B. *Rec. Trav. Chim. Pays Bas* **1925**, *44*, 141.
8. Trofimov, B. A.; Vasiltssov, A. M.; Mikhaleva, A. I.; Kalabin, G. A.; Shcherbakov, V. V.; Nesterenko, R. N.; Polubentsev, E. A.; Praliev, K. D. *Khim. Geterotsikl. Soedin.* **1991**, 1365.
9. Aliev, A. E.; Sinitsyna, A. A.; Borisova, T. N.; Stazharova, I. A.; Prostakov, N. S.; Varlamov, A. V. *Chem. Heterocycl. Compd. (Engl. Transl.)* **1993**, *29*, 65.
10. Quintana, R. P.; Lasslo, A.; Dugdale, M.; Goodin, L. L. *Thrombosis Res.* **1981**, *22*, 665.
11. Lawrence, W. H.; Howell, R. D.; Gollamudi, R. *J. Pharm. Sci.* **1994**, *83*, 222.
12. Maclog P 3.0 program (BioByte Corp., Claremont, CA, USA).
13. Hansch, C.; Leo, A. *Substituent Constants for Correlation Analysis in Chemistry and Biology*; John Wiley & Sons: New York, 1979.
14. Leo, A.; Hansch, C.; Elkins, D. *Chem. Rev.* **1971**, *71*, 525.
15. A LUNA 5 $\mu$  C18 column (150 $\times$ 4.6 mm i.d.) (Phenomenex, Torrance, CA, USA) was used as the stationary phase. The mobile phases consisted of different volume fractions of methanol in 0.01 M phosphate buffer (pH 4.50). All the measurements were made at a constant temperature of 25 $\pm$ 0.5 $^{\circ}$ C and flow-rate of 1.0 mL/min on a Waters HPLC Model 600 multisolvent delivery system (Waters Assoc., Milford, MA, USA).
16. Altomare, C.; Cellamare, S.; Carotti, A.; Ferappi, M. *Quant. Struct-Act. Relat.* **1993**, *12*, 261.
17. Altomare, C.; Cellamare, S.; Carotti, A.; Ferappi, M. *Farmaco* **1994**, *49*, 393.
18. Minick, D. J.; Brent, D. A.; Frenz, J. *J. Chromatogr. (A)* **1989**, *461*, 177.
19. Chen, N.; Zhang, Y.; Lu, P. *J. Chromatogr. (A)* **1993**, *633*, 31.
20. Luco, J. M.; Yamin, L. J.; Ferretti, H. F. *J. Pharm. Sci.* **1995**, *84*, 903.
21. Abraham, M. H.; Chadha, H. S.; Mitchell, R. C. *J. Pharm. Sci.* **1994**, *83*, 1257.
22. Boyd, M.; Boyd, P. D. W.; Atwell, G. J.; Wilson, W. R.; Denny, W. A. *J. Chem. Soc. Perkin Trans.* **1992**, *2*, 579.
23. Feng, Z.; Gollamudi, R.; Dillingham, E. O.; Bond, S. E.; Lyman, B. A.; Purcell, W. P.; Hill, R. J.; Korfmacher, W. A. *J. Med. Chem.* **1992**, *35*, 2952.